

## Recent progress in intrasynovial flexor tendon repair and rehabilitation

M.I. Boyer

Department of Orthopaedic Surgery, Washington University, St. Louis, MO, USA

**Keywords:** Tendon, Canine, Rehabilitation, Excursion, Force

Significant advances in the understanding of intrasynovial flexor tendon repair and rehabilitation have been made since the early 1970s<sup>1,2</sup> when reports first demonstrated that flexor tendon lacerations within the fibroosseous digital sheath could be repaired primarily and rehabilitated successfully without tendon excision and delayed grafting<sup>3</sup>. The concept of adhesion-free, or intrinsic tendon healing – that tendons could heal primarily without the ingrowth of fibrous adhesions from the surrounding sheath has been validated both experimentally and clinically in studies over the past 20 years<sup>4-13</sup>. Recent attempts to understand and improve the results of intrasynovial flexor tendon repair have focused upon restoration of the gliding surface<sup>11,14-23</sup>, repair site biomechanics<sup>24-37</sup> and on the molecular biology of early tendon healing<sup>38-47</sup>. The goals of the surgical treatment of intrasynovial flexor tendon lacerations remain unchanged, however: to achieve a primary tendon repair of sufficient tensile strength to allow application of a post-operative passive motion rehabilitation protocol such that formation of intrasynovial adhesions is inhibited, restoration of the gliding surface is stimulated, and healing of the repair site is facilitated<sup>48</sup>. I will focus upon two studies published recently from our tendon group at Washington University in St Louis that have evaluated intrasynovial repair site excursion and proximal musculotendinous load as independent variables, with the objective of determining whether or not rehabilitation programs that emphasize active motion are biologically advantageous to the healing tendon.

In an effort to improve the results of intrasynovial flexor tendon repair, authors have advocated rehabilitation methods that generate increased levels of applied *in vivo* force

and tendon excursion<sup>2,5,49,50</sup>. The clinical success of early passive motion rehabilitation has encouraged surgeons to prescribe rehabilitation protocols that further increase the "motion stress" on the repair site in order to stimulate healing<sup>48</sup>. However, there are no scientific data to support the concept that more aggressive rehabilitation accelerates tendon healing.

To address the issue of rehabilitation, we quantified, in an *in vivo* canine study, the flexor digitorum profundus (FDP) tendon force and excursion produced by five clinically relevant passive motion protocols<sup>51,52</sup>. Force was measured using a Z-transducer placed on the FDP tendon and excursion was assessed using a video technique. The combined excursion and force data indicate three distinct mechanical paradigms ( $p < 0.05$ ). First, with the wrist flexed and either one or four digits flexed and extended (1F, 4F), low excursion (1.7 mm) and low force (5 N) were produced. Second, with synergistic flexion of the digits as the wrist was extended (SYN), high excursion (3.5 mm) but low force were produced. Third, with the wrist and digits flexed and extended simultaneously (1E, 4E), high excursion and high force (17 N) were produced. These data described, for the first time, the combined excursion and force variables that operate on the canine FDP tendon during joint manipulations and allowed a set of experiments to be conducted in which excursion and force were varied independently of each other.

We first examined the effects of increased *in vivo* tendon excursion on digital range of motion and tendon strength following FDP tendon transection and repair<sup>53</sup>. Ninety-six FDP tendons from 48 dogs were injured and repaired. The affected forelimbs were treated by passive mobilization during two five-minute rehabilitation sessions, performed five days a week starting on the first post-operative day. Rehabilitation for one group (4F; low-force/low-excision) was performed after removing the volar flexion block and consisted of passive flexion and extension of the four digits with the wrist maintained in the flexed position. For the second group (4E, low-force/high-excision) both the volar flexion and dorsal extension blocks were removed and the wrist and digits flexed and extended simultaneously. Both protocols were

The author has no conflict of interest.

Corresponding author: Martin I. Boyer, M.D., Assistant Professor, Department of Orthopaedic Surgery, Washington University, St Louis, One Barnes Plaza, Suite 11300, St. Louis, MO 63110, USA  
E-mail: boyerm@msnotes.wustl.edu

Accepted 1 August 2003

performed at a rate of approximately 1 Hz, resulting in 600 cycles of loading each day. Dogs received rehabilitation daily until sacrifice at 10, 21 or 42 days.

Our results indicated that the use of rehabilitation that produced increased tendon excursion did not influence range-of-motion or tensile properties. Joint rotation and tendon excursion in digits from the low-force/low-excision and low-force/high-excision groups were not significantly different ( $p > 0.05$ ), with both groups not different from unoperated controls. Tensile structural properties (ultimate force, rigidity, strain at 20 N, strain at failure) were not significantly affected by increased excursion ( $p > 0.05$ ). We conclude that a threshold of 1.7 mm of tendon excursion is sufficient to inhibit adhesion formation and to allow excellent recovery of functional properties following sharp transection of the canine FDP tendon. Additional excursion, at the same low force level, provides little added benefit.

The objective of our next study was to assess the effects of variations in applied *in vivo* force on biomechanical properties. We compared low-force/high-excision versus high-force/high-excision rehabilitation, using 246 tendons from 123 dogs<sup>25</sup>. Because we hypothesized that there might be an interaction between the stiffness of the repair and the level of applied rehabilitation force, we used both 4- and 8-strand repairs in this study.

Our results indicated that tensile properties were not different between low- and high-force rehabilitation groups, regardless of repair technique. Rehabilitation method did not significantly affect ultimate force ( $p=0.48$ ), repair-site rigidity ( $p=0.96$ ), strain at 20 N ( $p=0.29$ ) or strain at failure ( $p=0.22$ ). For example, in the 42-day, 4-strand group, tendons treated with high-force rehabilitation had an average ultimate force of 96 N, which was not significantly different from the average of 102 N for the low-force rehabilitation group. Moreover, method of rehabilitation had no significant effect on distal or proximal interphalangeal joint flexion (distal,  $p = 0.91$ ; proximal,  $p = 0.87$ ) and only a slight effect on tendon displacement (9% less in the high-force group;  $p = 0.024$ ). Operated digits had approximately the same range of motion as contralateral control digits, with proximal interphalangeal joint flexion ( $p = 0.20$ ) and tendon displacement ( $p = 0.58$ ) not significantly different and distal interphalangeal joint flexion reduced by only 11% compared to control ( $p < 0.001$ ).

Based on these results, we conclude that increasing the level of force within the range that can be applied using passive mobilization (i.e., 5 to 17 N), does not accelerate the time-dependent accrual of stiffness and strength in the canine model. Taken together with the previous finding that increased tendon excursion did not enhance tendon healing<sup>53</sup>, our findings suggest that there be a re-examination of the widely held concept that increases in force and motion produced by more vigorous mobilization protocols are beneficial to tendon healing. While more vigorous rehabilitation may help improve overall hand function, we found no evidence that it enhances tissue healing and strength in the context of a modern suture repair.

Several conclusions can be drawn from our recent findings using the clean laceration canine model with multistrand suture repair and early passive motion rehabilitation. First, suture technique is of primary importance in providing a stiff and strong repair throughout the early healing interval, and the benefits of a multistrand repair are observed regardless of the level of rehabilitation force. Second, increased tendon excursion beyond the amount produced by passive digital flexion-extension (with the wrist flexed) does not enhance digital motion or tendon healing. Third, application of increased levels of passive force during the early post-operative period is not supported as a means to accelerate tendon healing, since the time-dependent accrual of stiffness and strength does not appear to be enhanced by increased force levels within a clinically relevant range. In order to increase the sensitivity of the repair site to increased force levels, it may be necessary to alter the biochemical environment to increase expression of integrins or other force-sensitive molecules. Future strategies to accelerate tissue healing may therefore require manipulation of both biochemical and rehabilitation variables.

## References

1. Verdan CE. Half a century of flexor-tendon surgery. Current status and changing philosophies. *J Bone Joint Surg Am* 1972; 54:472-491.
2. Kessler I, Nissim F. Primary repair without immobilization of flexor tendon division within the digital sheath. An experimental and clinical study. *Acta Orthop Scand* 1969; 40:587-601.
3. Bunnell S. *Surgery of the Hand*. Edited, 381-466, Lippincott, Philadelphia; 1948.
4. Gelberman RH, Amiel D, Gonsalves M, Woo S, Akeson WH. The influence of protected passive mobilization on the healing of flexor tendons: a biochemical and microangiographic study. *Hand* 1981; 13:120-128.
5. Gelberman RH, Woo SL, Lothringer K, Akeson WH, Amiel D. Effects of early intermittent passive mobilization on healing canine flexor tendons. *J Hand Surg [Am]* 1982; 7:170-175.
6. Manske PR, Gelberman RH, Vande Berg JS, Lesker PA. Intrinsic flexor-tendon repair. A morphological study *in vitro*. *J Bone Joint Surg Am* 1984; 66:385-396.
7. Manske PR, Lesker PA. Biochemical evidence of flexor tendon participation in the repair process—an *in vitro* study. *J Hand Surg [Br]* 1984; 9:117-120.
8. Manske PR, Lesker PA. Histologic evidence of intrinsic flexor tendon repair in various experimental animals. An *in vitro* study. *Clin Orthop* 1984; (182):297-304.
9. Gelberman RH, Manske PR, Vande Berg JS, Lesker PA, Akeson WH. Flexor tendon repair *in vitro*: a comparative histologic study of the rabbit, chicken, dog, and monkey. *J Orthop Res* 1984; 2:39-48.
10. Gelberman RH, Vandeberg JS, Manske PR, Akeson

- WH. The early stages of flexor tendon healing: a morphologic study of the first fourteen days. *J Hand Surg [Am]* 1985; 10:776-784.
11. Manske PR, Lesker PA, Gelberman RH, Rucinsky TE. Intrinsic restoration of the flexor tendon surface in the nonhuman primate. *J Hand Surg [Am]* 1985; 10:632-637.
  12. Manske PR, Gelberman RH, Lesker PA. Flexor tendon healing. *Hand Clin* 1985; 1:25-34.
  13. Gelberman RH, Manske PR, Akeson WH, Woo SL, Lundborg G, Amiel D. Flexor tendon repair. *J Orthop Res* 1986; 4:119-128.
  14. Zhao C, Amadio PC, Zobitz ME, Momose T, Couvreur P, An K. Gliding resistance after repair of partially lacerated human flexor digitorum profundus tendon *in vitro*. *Clin Biomech (Bristol, Avon)* 2001; 16:696-701.
  15. Zhao C, Amadio PC, Zobitz ME, An KN. Gliding characteristics of tendon repair in canine flexor digitorum profundus tendons. *J Orthop Res* 2001; 19:580-586.
  16. Momose T, Amadio PC, Zhao C, Zobitz ME, An KN. The effect of knot location, suture material, and suture size on the gliding resistance of flexor tendons. *J Biomed Mater Res* 2000; 53:806-811.
  17. Boardman ND III, Morifusa S, Saw SS, McCarthy DM, Sotereanos DG, Woo SL. Effects of tenorrhaphy on the gliding function and tensile properties of partially lacerated canine digital flexor tendons. *J Hand Surg [Am]* 1999; 24:302-309.
  18. Aoki M, Kubota H, Pruitt DL, Manske PR. Biomechanical and histologic characteristics of canine flexor tendon repair using early post-operative mobilization. *J Hand Surg [Am]* 1997; 22:107-114.
  19. Gelberman RH, Vande Berg JS, Lundborg GN, Akeson WH. Flexor tendon healing and restoration of the gliding surface. An ultrastructural study in dogs. *J Bone Joint Surg Am* 1983; 65:70-80.
  20. Nyska M, Porat S, Nyska A, Rouso M, Shoshan S. Decreased adhesion formation in flexor tendons by topical application of enriched collagen solution – a histological study. *Arch Orthop Trauma Surg* 1987; 106:192-194.
  21. Porat S, Rouso M, Shoshan S. Improvement of gliding function of flexor tendons by topically applied enriched collagen solution. *J Bone Joint Surg Br* 1980; 62-B(2):208-213.
  22. Peterson WW, Manske PR, Kain CC, Lesker PA. Effect of flexor sheath integrity on tendon gliding: a biomechanical and histologic study. *J Orthop Res* 1986; 4:458-465.
  23. Zhao C, Amadio PC, An KN, Zobitz ME. Gliding characteristics of tendon repair in partially lacerated canine flexor digitorum profundus tendons. In 45th Ann Orthop Res Soc (Edited), Anaheim; 1999:120.
  24. Boyer MI, Meunier MJ, Lescheid J, Burns ME, Gelberman RH, Silva MJ. The influence of cross-sectional area on the tensile properties of flexor tendons. *J Hand Surg [Am]* 2001; 26:828-832.
  25. Boyer MI, Gelberman RH, Burns ME, Dinopoulos H, Hofem R, Silva MJ. Intrasynovial flexor tendon repair. An experimental study comparing low and high levels of *in vivo* force during rehabilitation in canines. *J Bone Joint Surg Am* 2001; 83-A:891-899.
  26. Miller L, Mass DP. A comparison of four repair techniques for Camper's chiasma flexor digitorum superficialis lacerations: tested in an *in vitro* model. *J Hand Surg [Am]* 2000; 25:1122-1126.
  27. Dinopoulos HT, Boyer MI, Burns ME, Gelberman RH, Silva MJ. The resistance of a four- and eight-strand suture technique to gap formation during tensile testing: an experimental study of repaired canine flexor tendons after 10 days of *in vivo* healing. *J Hand Surg [Am]* 2000; 25:489-498.
  28. Choueka J, Heminger H, Mass DP. Cyclical testing of zone II flexor tendon repairs. *J Hand Surg [Am]* 2000; 25:1127-1134.
  29. Gelberman RH, Boyer MI, Brodt MD, Winters SC, Silva MJ. The effect of gap formation at the repair site on the strength and excursion of intrasynovial flexor tendons. An experimental study on the early stages of tendon-healing in dogs. *J Bone Joint Surg Am* 1999; 81:975-982.
  30. Winters SC, Gelberman RH, Woo SL-Y, Chan SS, Grewal R, Seiler JG. The effects of multiple-strand suture methods on the strength and excursion of repaired intrasynovial flexor tendon: a biomechanical study in dogs. *J Hand Surg [Am]* 1998; 23:97-104.
  31. Stein T, Ali A, Hamman J, Mass DP. A randomized biomechanical study of zone II human flexor tendon repairs analyzed in an *in vitro* model. *J Hand Surg [Am]* 1998; 23:1046-1051.
  32. Silva MJ, Hollstien SB, Fayazi AH, Adler P, Gelberman RH, Boyer MI. The effects of multiple-strand techniques on the tensile properties of flexor digitorum profundus to bone repair. *J Bone Jt Surg [Am]* 1998; 80:1507-1514.
  33. Winters SC, Seiler JG III, Woo SL, Gelberman RH. Suture methods for flexor tendon repair. A biomechanical analysis during the first six weeks following repair. *Ann Chir Main Memb Super* 1997; 16:229-234.
  34. Hamman J, Ali A, Phillips C, Cunningham B, Mass DP. A biomechanical study of the flexor digitorum superficialis: effects of digital pulley excision and loss of the flexor digitorum profundus. *J Hand Surg [Am]* 1997; 22:328-335.
  35. Komanduri M, Phillips CS, Mass DP. Tensile strength of flexor tendon repairs in a dynamic cadaver model. *J Hand Surg [Am]* 1996; 21:605-611.
  36. Noguchi M, Seiler JG III, Gelberman RH, Sofranko RA, Woo SL. *In vitro* biomechanical analysis of suture methods for flexor tendon repair. *J Orthop Res* 1993; 11:603-611.
  37. Goomer RS, Maris TM, Gelberman R, Boyer M, Silva

- M, Amiel D. Nonviral *in vivo* gene therapy for tissue engineering of articular cartilage and tendon repair. Clin Orthop 2000; 379(Suppl.):S189-200.
38. Bidder M, Towler DA, Gelberman RH, Boyer MI. Expression of mRNA for vascular endothelial growth factor at the repair site of healing canine flexor tendon. J Orthop Res 2000; 18:247-252.
  39. Tsuzaki M, Brigman BE, Yamamoto J, Lawrence WT, Simmons JG, Mohapatra NK, Lund PK, Van Wyk J, Hannafin J, Bhargava MM, Banes AJ. Insulin-like growth factor-I is expressed by avian flexor tendon cells. J Orthop Res 2000; 18:546-556.
  40. Banes AJ, Weinhold P, Yang X, Tsuzaki M, Bynum D, Bottlang M, Brown T. Gap junctions regulate responses of tendon cells *ex vivo* to mechanical loading. Clin Orthop 1999; 367(Suppl.):S356-370.
  41. Banes AJ, Horesovsky G, Larson C, Tsuzaki M, Judex S, Archambault J, Zernicke R, Herzog W, Kelley S, Miller L. Mechanical load stimulates expression of novel genes *in vivo* and *in vitro* in avian flexor tendon cells. Osteoarthritis Cartilage 1999; 7:141-153.
  42. Banes AJ, Tsuzaki M, Hu P, Brigman B, Brown T, Almekinders L, Lawrence WT, Fischer T. PDGF-BB, IGF-I and mechanical load stimulate DNA synthesis in avian tendon fibroblasts *in vitro*. J Biomech 1995; 28:1505-1513.
  43. Chang J, Thunder R, Most D, Longaker MT, Lineaweaver WC. Studies in flexor tendon wound healing: neutralizing antibody to TGF- $\beta$ 1 increases post-operative range of motion. Plast Reconstr Surg 2000; 105:148-155.
  44. Chang J, Most D, Thunder R, Mehrara B, Longaker MT, Lineaweaver WC. Molecular studies in flexor tendon wound healing: the role of basic fibroblast growth factor gene expression. J Hand Surg [Am] 1998; 23:1052-1058.
  45. Chang J, Most D, Stelnicki E, Siebert JW, Longaker MT, Hui K, Lineaweaver WC. Gene expression of transforming growth factor beta-1 in rabbit zone II flexor tendon wound healing: evidence for dual mechanisms of repair. Plast Reconstr Surg 1997; 100(4):937-944.
  46. Boyer MI, Watson JT, Lou J, Manske PR, Gelberman RH, Cai SR. Quantitative variation in vascular endothelial growth factor mRNA expression during early flexor tendon healing: an investigation in a canine model. J Orthop Res 2001; 19:869-872.
  47. Strickland JW. Development of flexor tendon surgery: twenty-five years of progress. J Hand Surg [Am] 2000; 25:214-235.
  48. Lister GD, Kleinert HE, Kutz JE, Atasoy E. Primary flexor tendon repair followed by immediate controlled mobilization. J Hand Surg [Am] 1977; 2:441-451.
  49. Strickland JW, Glogovac SV. Digital function following flexor tendon repair in Zone II: a comparison of immobilization and controlled passive motion techniques. J Hand Surg [Am] 1980; 5:537-543.
  50. Lieber RL, Amiel D, Kaufman KR, Whitney J, Gelberman RH. Relationship between joint motion and flexor tendon force in the canine forelimb. J Hand Surg [Am] 1996; 21: 957-962.
  51. Lieber RL, Silva MJ, Amiel D, Gelberman RH. Wrist and digital joint motion produce unique flexor tendon force and excursion in the canine forelimb. J Biomech 1999; 32:175-181.
  52. Silva MJ, Brodt MD, Boyer MI, Morris TS, Dinopoulos H, Amiel D, Gelberman RH. Effects of increased *in vivo* excursion on digital range of motion and tendon strength following flexor tendon repair. J Orthop Res 1999; 17:777-783.